# SUBTILISIN-CATALYZED HYDROLYSIS OF PEPTIDE METHYL ESTERS

Manfred PHILIPP and Myron L. BENDER

Division of Biochemistry, Department of Chemistry, Northwestern University, Evanston, Illinois 60201, USA

Received 5 February 1974

#### 1. Introduction

It is known that the serine protease subtilisin possesses a specifity for longer peptide amides as opposed to simpler amides [1,2]. Since ester substrates of the serine protease chymotrypsin generally are hydrolyzed much more rapidly ( $10^5$  times) [3] than amides, the subtilisin-catalyzed hydrolyses of specific peptide esters should be much more rapid than those previously studied. Two substrates used here have the highest known reaction rate  $(k_2/K_s)$  with subtilisin. These rates, for methyl esters, approximate the rates of the best substrates of  $\alpha$ -chymotrypsin [4] which are more highly activated p-nitrophenyl esters.

Since in the most specific substrates, the value of  $k_2$  may exceed  $k_1$  (eq. 1), the value of  $k_{\rm cat}/K_{\rm m}$  may reduce to  $k_1$ . [4]

$$E + S \stackrel{k_1}{=} ES \stackrel{k_2}{\longleftrightarrow} EP_2 \stackrel{k_3}{\to} E + P_2$$
 (1)

It is evident that the pH profile of  $k_1$  need not be identical to that of  $k_2/K_s$ . [5] Therefore pH profiles of the various hydrolyses were obtained. Since literature data [1,2] concerning the peptide amides similar to the esters is in the form of relative rate data, binding constants of the amides to subtilisin were obtained. These were measured by observing inhibition of the second-order subtilisin-catalyzed hydrolysis of Z-Gly-OnP (see later).

# 2. Materials and methods

Subtilisin Novo was purchased from Novo Industri, Copenhagen, Denmark. Its molar concentration was

determined using a cinnamov limidazole titration [6]. All substrates and inhibitors were purchased from the Cyclo Chemical Company. p-Nitrophenyl N-benzyloxycarbonyl glycinate (Z-Gly-OnP) [7] was purified by dissolving it twice in acetone and adding heptane until the onset of precipitation. This was followed by refrigeration. The hydrolysis of this compound was observed at 400 nm, of the tyrosine methyl ester at 237 nm [8], and of the tryptophan methyl ester at 300 nm [9]. Concentrations of Z-Gly-Gly-L-Tyr-OMe used varied from 3 to  $10 \times 10^{-5}$  M, those of Z-Gly-Gly-L-Trp-OMe from 2 to  $4 \times 10^{-4}$  M, and those of Ac-L-Trp-OMe from 3 to  $4 \times 10^{-4}$  M were used in pH dependence studies, and organic solvent concentrations (methanol or DMSO) were 2.5%. Inhibition studies were performed at pH 7.71 I = 0.1 M phosphate buffer [10] with 4.4% N, N-dimethylformamide. Inhibitor concentrations ranged from 2 to  $20 \times 10^{-3}$  M. Binding constants were calculated using an expression used previously. [11]

#### 3. Results and discussion

## 3.1. Hydrolysis of methyl ester substrates

Two parameters, the p $K_1$  of  $k_2/K_s$  and  $k_2/K_s$  (lim) were measured for Ac-Ty-OMe, Z-Gly-Gly-Try-OMe and Z-Gly-Gly-Tyr-OMe. Figs. 1–3 show the pH profiles and the profile parameters for these substrates. Comparison of the parameters for Ac-Try-OMe and Z-Gly-Gly-Try-OMe shows that the increase in the second-order acylation rate constant  $k_2/K_s$  (lim) is not accompanied by any significant change in pK.

The  $k_2/K_s$  (lim) value for Z-Gly-Gly-Tyr-OMe (2.6 × 10<sup>7</sup> M<sup>-1</sup> sec<sup>-1</sup>) is the highest yet observed for

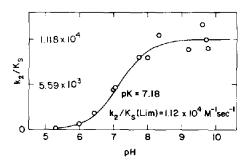


Fig. 1. pH Dependence of the subtilisin-catalyzed hydrolysis of methyl N-acetyl-tryptophanate. pK = 7.18,  $k_2/K_8$  (lim) =  $1.12 \times 10^4~{\rm M}^{-1}~{\rm sec}^{-1}$ .

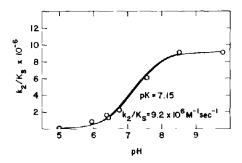


Fig. 2. pH dependence of the subtilisin-catalyzed hydrolysis of methyl N-CBZ-glycyl-glycyl-L-tryptophanate. pK = 7.15,  $k_2/K_s$  (lim) =  $9.2 \times 10^6 \text{ M}^{-1} \text{ sec}^{-1}$ .

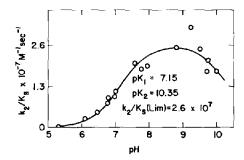


Fig. 3. pH Dependence of the subtilisin-catalyzed hydrolysis of methyl N-CBZ-glycyl-glycyl-L-tyrosinate. p $K_1 = 7.15$ , p $K_2 = 10.35$ ,  $k_2/K_S$  (lim) =  $2.6 \times 10^7 \ \mathrm{M}^{-1} \ \mathrm{sec}^{-1}$ .

a subtilisin substrate. The literature value for Ac-Tyr-OMe is  $1.7 \times 10^4 \text{ M}^{-1}\text{sec}^{-1}$ . [12] Hydrolysis of Z-Gly-Gly-Tyr-OMe also shows the pK of 7.15 observed for Ac-Try-OMe.

Very high  $k_2/K_s$  values may be associated with rate determining binding,  $k_1$  (eq. 1). Since  $K_m$  for subtilisin substrates is independent of pH in the region 6–9, [1,13] the individual constants,  $k_1$  and  $k_{-1}$ , are likely to be pH independent in this region. Thus, if  $k_1$  becomes rate determining, the pH profile should be perturbed. [5] This evidently does not happen, and so  $k_1$  for these substrates must be considerably higher than  $2.6 \times 10^7 \, \mathrm{M}^{-1} \, \mathrm{sec}^{-1}$ .

The magnitude of the ester rates observed here may also be compared to the magnitude of  $k_2/K_8$  (lim) for the best amide substrate known for subtilisin, Z-Gly Ala-Leu-NH<sub>2</sub>, which is  $6.3 \times 10^3$  M<sup>-1</sup> sec<sup>-1</sup>. [1]

Thus, even the most highly specific amide substrates are still hydrolyzed much more slowly  $(10^3 - 10^4)$  than similar ester substrates.

Finally, it is apparent that Z-Gly-Gly-Tyr-OMe shows a  $k_2/K_8$  pK near 10.3. Since the subtilisin-catalyzed hydrolysis of Bz-Tyr-OEt also shows this pK [14] while the analogous tryptophan substrates do not, it is reasonable to conclude that the substrate tyrosine phenolic group ionization causes this pK.

#### 3.2. Inhibition constants of peptide amides

Table 1 shows binding constants of various peptide amides, measured by observing inhibition of the second-order hydrolysis of Z-Gly-OnP. In each series the binding constant is strongly determined by the presence or absence of the N-benzyloxycarbonyl group, which is known to bind as a phenylalanine side-chain [15]. This effect is nearly independent of the peptide chain length. Crystal structure studies [15] indicate that nonproductive binding modes are

Table 1

Inhibitor	$K_{\mathbf{I}}$ (M)
Ac-Tyt-NH <sub>2</sub>	1.2 × 10 <sup>-1</sup>
Z-Tyr-NH,	$7.7 \times 10^{-3}$
Z-Gly-Tyr-NH <sub>2</sub>	$8.6 \times 10^{-3}$
Z-Gly-Gly-Tyr-NH,	$5.1 \times 10^{-3}$
Z-Gly-Trp-NH,	$7.5 \times 10^{-3}$
Z-Gly-Gly-Trp-NH,	$3.6 \times 10^{-3}$

present in these complexes which can affect  $k_{\text{cat}}$ ,  $K_{\text{m}}$ , and  $K_{\text{l}}$  values in a similar way.

If the  $K_{\rm I}$  values are taken to be similar to the  $K_{\rm m}$  value (which includes inhibitory nonproductive components) estimates of  $k_2$  for Z-Gly-Gly-Tyr-OMe and Ac-Tyr-OMe can be made. These are  $1\times10^4$  sec<sup>-1</sup> for Z-Gly-Gly-Tyr-OMe and  $2\times10^3$  sec<sup>-1</sup> for Ac-Tyr-OMe. These values are calculated by substituting the value of  $K_{\rm I}$  for  $K_{\rm S}$  in the value of  $k_2/K_{\rm S}$ . These values are quite similar to those of specific chymotrypsin substrates obtained by similar indirect and direct means.  $k_2$  For the chymotrypsin-catalyzed hydrolysis of Ac-Tyr-OEt is  $5.33\times10^3$  sec<sup>-1</sup> [16] similar to the estimated subtilisin value with the methyl ester.

## 4. Acknowledgement

This research was supported by grant HLO-5726 of the U.S. National Institutes of Health.

#### References

[1] Morihara, K., Oka, T., and Tsuzuki, H. (1970) Arch. Biochem. Biophys. 138, 515.

- [2] Morihara, K., Tsuzuki, H., and Oka, T. (1971) Biochem. Biophys. Res. Comm., 42, 1000.
- [3] Bender, M. L., and Kezdy, F. J. (1965) Ann. Rev. Biochem., 34, 66.
- [4] Philipp, M. and Bender, M. L. (1973) Nature, New Biology, 241, 44.
- [5] Renard, M. and Fersht, A. R., (1973) Biochemistry, 12, 4713.
- [6] Bender, M. L. et al. (1966) J. Amer. Chem. Soc., 88, 5890
- [7] Polgar, L. and Bender, M. L. (1969) Biochemistry, 8,
- [8] Schwert, G. W. and Takenaka, Y. (1955) Biochim. Biophys. Acta, 16, 570.
- [9] Zerner, B. and Bender, M. L. (1964) J. Amer. Chem. Soc., 86, 3669.
- [10] Long, C. (1966) ed. Biochemists' Handbook, D. Van Nostrand, Princeton, New Jersey, pp. 30-42.
- [11] Philipp, M. and Bender, M. L. (1971) Proc. Natl. Acad. Sci., U.S., 68, 478.
- [12] Barel, A. O. and Glazer, A. N. (1968) J. Biol. Chem., 243, 1844.
- [13] Glazer, A. N. (1969) J. Biol. Chem., 242, 433.
- [14] Gounaris, A. and Ottesen, M. (1966) C. R. Trav. Lab. Carlsberg, 35, 37.
- [15] Robertus, J. D., Kraut, J., Alden, R. A., and Birktoft, J. J. (1972) Biochemistry, 11, 4293.
- [16] McConn, J., Ku, E., Himoe, A., Brandt, K. G. and Hess, G. P. (1971) J. Biol. Chem., 246, 2918.